CORRECTIONS

Spin-Label Detection of Hemoglobin–Membrane Interaction at Physiological pH, by Leslie W.-M. Fung, Volume 20, Number 25, December 8, 1981, pages 7162–7166.

Pages 7163 and 7164. In the legends to Figures 1 and 2, the plot symbols should be as follows: pH 8.0 (■), 7.4 (▲), and 6.7 (●). The top panels in Figures 1 and 2 show the data of pH 8.0 and the bottom panels show the data of pH 6.7.

Purification and Physicochemical Properties of Starch Phosphorylase from Young Banana Leaves, by Anil Kumar and G. G. Sanwal*, Volume 21, Number 17, August 17, 1982, pages 4152–4159.

Page 4156. In Figure 5, line b in the graph should be read as line c and vice versa.

Reduction, Oxidation, and Addition Reactions between Free Radicals and Flavins, by Rizwan Ahmad and David A. Armstrong*, Volume 21, Number 22, October 26, 1982, pages 5445–5450.

The extinction coefficient of 6000 for lumichrome (LC) in Figure 4b was based on data given by Koziol [(1966) *Photochem. Photobiol. 5*, 41–54]. While other flavin extinction coefficients that were taken from the literature and used for determining flavin concentrations are correct, a redetermination in our laboratory gave ϵ (LC at 350 nm) = 9830 M^{-1} cm^{-1} (±2%) at pH 7. The ordinate scale on Figure 4b should therefore be changed by a factor of 1.64 as should the values of η, the uptake of reducing or oxidizing equivalents of radical per mole of LC in titrations. The average value of η for reduction by $\cdot CO_2^-$ is thus changed from 1.1 to 1.8, which implies that the major product is a dihydroalloxazine. On the basis of stoichiometry, the dimer (i.e., HLC–LCH), which was postulated in our penultimate paragraph to explain η near unity, is now seen to make up only a fraction (about 25%) of the reduced product. The recalculated value of η = 1.6 for $\cdot Br_2^-$ oxidation of this product supports this. Further work will be published elsewhere.

Stoichiometry of the H^+-ATPase of Growing and Resting, Aerobic *Escherichia coli*, by Eva R. Kashket, Volume 21, Number 22, October 26, 1982, pages 5534–5538.

Page 5535. In column 2, under Determination of Cellular Adenine Nucleotide Concentrations, line 4 should read as follows: 0.5 mL of ice-cold 90% trichloroacetic acid.

Interactions between Phospholipid Head Groups at Membrane Interfaces: A Deuterium and Phosphorus Nuclear Magnetic Resonance and Spin-Label Electron Spin Resonance Study, by Frank Sixl and Anthony Watts*, Volume 21, Number 25, December 7, 1982, pages 6446–6452.

Page 6447. In line 2 under Materials and Methods, 1,2-dimyristoyl-*sn*-glycero-3-phospho-*rac*-glycerol should read 1,2-dimyristoyl-*sn*-glycero-3-phospho-*sn*-glycerol.

Analysis of the Rate-Limiting Step in a Ligand–Cell Receptor Interaction: The Immunoglobulin E System, by Stephen A. Wank, Charles DeLisi, and Henry Metzger*, Volume 22, Number 4, February 15, 1983, pages 954–959.

Page 957. In Table III, the units of k_f should be M^{-1} s^{-1} × 10^{-5}.

Reversible Inhibition of the Bacterial Luciferase Catalyzed Bioluminescence Reaction by Aldehyde Substrate: Kinetic Mechanism and Ligand Effects, by Thomas F. Holzman and Thomas O. Baldwin*, Volume 22, Number 12, June 7, 1983, pages 2838–2846.

Page 2840. In column 2, the equation on line 13 of the text should read as follows: $\alpha = 4[v_0/(V_m - v_0)]^2$.

Page 2842. In column 2, the expression in parentheses beginning on line 21 should read (for EAP → EA_2P vs. EA → EA_2).

Page 2843. In the caption to Figure 6, the apparent kinetic and equilibrium constants for *n*-dodecanal should be K_m = 0.12 μM and $K_{i,app}$ = 9.3 μM. In Table III, the values for *n*-octanal should read α = 21, K_d = 8.8 μM, and αK_d (K_i) = 180 μM.

Rotational Diffusion of *Escherichia coli* RNA Polymerase Free and Bound to Deoxyribonucleic Acid in Nonspecific Complexes, by R. H. Austin, Joanne Karohl, and Thomas M. Jovin*, Volume 22, Number 13, June 21, 1983, pages 3082–3090.

Page 3084. In eq 5, the latter part should read as follows: $D_s = kT/(6V_h\eta)$.